Food Flour Proteins with Defined Allergen Composition for Use as Reference Materials #111 Catherine Thorpe, Lisa Vailes, Sayeh Agah, Martin Chapman, Sabina Wünschmann

RATIONALE

Allergen measurements are widely used for validation of molecular allergy diagnostics and allergy therapeutics. However, few standardized food allergen reference materials have been developed. While NIST and MoniQA food standards are characterized extensively for biochemical and nutritional composition, data on allergen content are lacking. The aim was to produce standardized food-flour-proteins with defined allergen content that could serve as reference materials for allergy diagnostics or therapeutics.

METHODS

Food-flour-proteins were prepared from defatted peanut-, hazelnut-, pistachio- and soyflour using optimized aseptic extraction conditions. Allergen composition was analyzed using validated allergen-specific ELISA's or mass spectrometry (LC-MS/MS). Real time stability data were collected from frozen allergens.

RESULTS

Peanut flour was extracted in PBS, 1M NaCl pH 7.4. Natural Ara h 1, Ara h 2, Ara h 3, and Ara h 6 were purified from blanched or roasted peanuts (Fig.1A). Peanut allergens Ara h 1 (220 µg/ml), Ara h 2 (371 µg/ml), Ara h 3 (879 µg/ml) and Ara h 6 (217 µg/ml) were quantified in peanut-flour-protein by ELISA in quadruplicate (Fig.1B). Endotoxin levels were < $0.03 \text{ EU/}\mu g$.

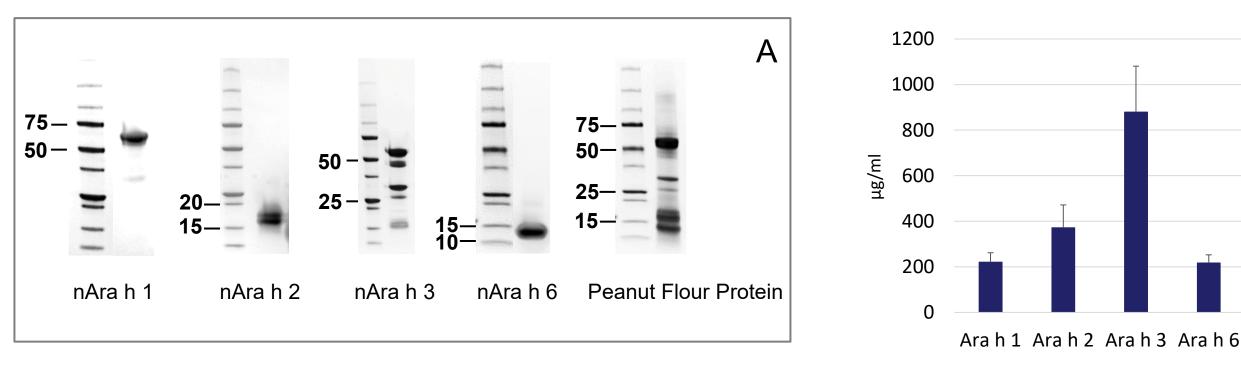


Fig. 1: A:SDS-PAGE of purified peanut allergens and peanut flour protein under non-reducing conditions followed by Coomassie Fig. 3: SDS-PAGE of purified Pis v 1 and Pistachio flour protein under non-reducing conditions followed by Coomassie staining. staining. B: Allergens in peanut food flour protein were quantified by peanut-allergen specific ELISA. Table 2: LC-MS/MS of allergens in pistachio flour protein

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RESULTS



Hazelnut flour was extracted in PBS, pH 7.4. Hazelnut allergens Cor a 1, Cor a 8, Cor a 9, Cor a 11, and Cor a 14 were purified from hazelnuts or expressed as recombinant proteins (Fig.2). LC-MS/MS analysis of hazelnut-flour confirmed the presence of Cor a 1, Cor a 8, Cor a 9, Cor a 11, Cor a 14 and Cor a 16 (Table 1).

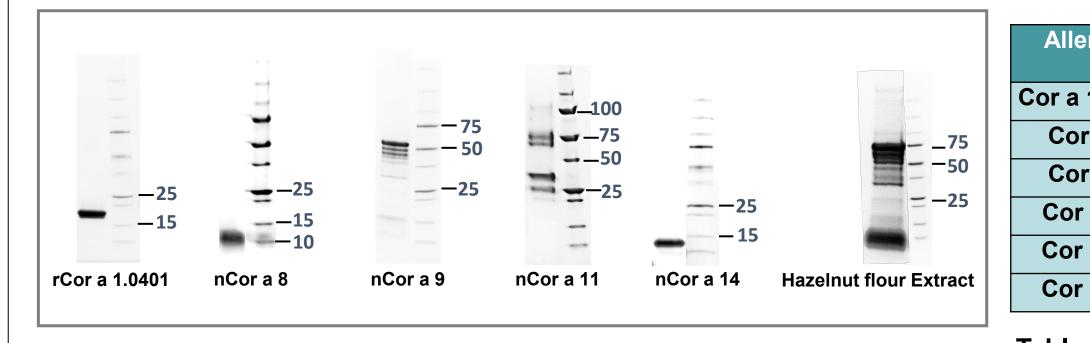
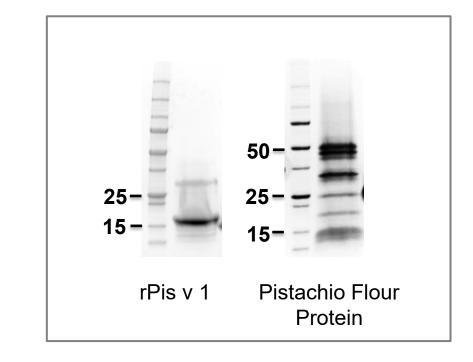




Fig. 2: SDS-PAGE of purified hazeInut allergens and hazeInut flour protein under non-reducing conditions followed by Coomassie staining. Table 1: LC-MS/MS of allergens in hazelnut flour protein.

Pistachio flour was extracted in 0.1M Potassium phosphate, 0.4M NaCI pH 8.0. Pistachio allergen Pis v 1 was expressed as recombinant protein (Fig.3). LC-MS/MS analysis of pistachio flour protein confirmed the presence of Pis v 1, Pis v 2, Pis v 3, Pis v 4, and Pis v 5 (Table 2).

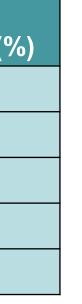


Allergen	Relative abundance (
Pis v 1	17%
Pis v 2	41%
Pis v 3	12%
Pis v 4	0.01%
Pis v 5	30%

Table 2*

RESULTS

ergen	Relative
	Abund. (%)
1.0401	0.2%
ra8	4%
ra9	51%
r a 11	13%
r a 14	15%
r a 16	17%



Soy flour was extracted in 50mM Tris, pH 8.2. Soy allergens Gly m 4, Gly m 5 and Gly m 6 were purified from soy or expressed as rec. proteins (Fig.4). LC-MS² analysis of soyflour confirmed the presence of Gly m 4, Gly m 5, Gly m 6, Gly m 7, and Gly m 8 (Table 3).

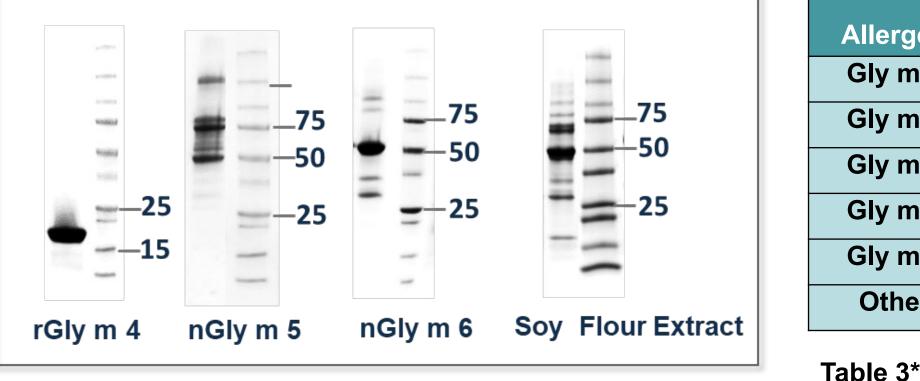
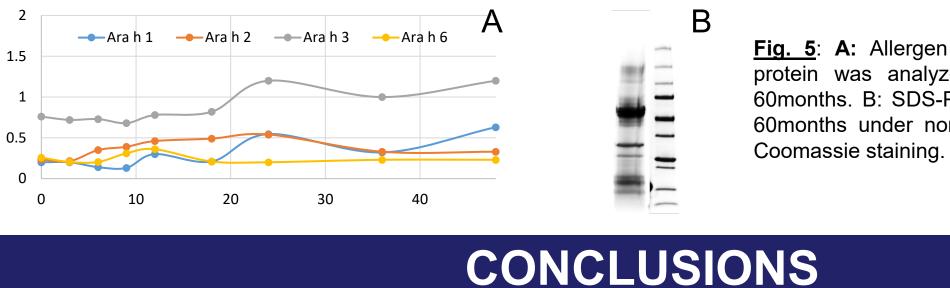


Fig. 4: SDS-PAGE of purified soy allergens and soy flour protein under non-reducing conditions followed by Coomassie staining. Table 3: LC-MS/MS of allergens in soy flour protein.

Stability Study of Peanut Flour Protein

Real time stability tests of frozen food-flour-proteins showed consistent protein content by ELISA and no signs of degradation on SDS-PAGE (Example peanut flour protein Fig. 5).



Optimized, ISO-9001 compliant, bioprocessing pathways have been established to yield standardized legume and tree nut food-flour-proteins with defined allergen profiles which can serve as food reference materials. The low-endotoxin, stable food-flour-proteins have applications as reference materials for monitoring the composition of allergy diagnostics and therapeutics.

*Note: (1) Relative abundance results are based on label-free quantification of precursor peptide ion intensity (peak area, incl. unique + razor peptides) using Proteome Discoverer 2.2 algorithms. (2) Calculations of relative abundance can be variable and depend on multiple factors such as sample complexity, buffer composition, instrument resolution and analysis software



	Relative
ergen	abundance (%)
/ m 4	0.03%
/ m 5	34%
/ m 6	46%
/ m 7	0.1%
/ m 8	2.5%
ther	17%

Fig. 5: A: Allergen composition of the peanut flour protein was analyzed by ELISA over a period of 60months. B: SDS-PAGE of peanut flour protein after 60months under non-reducing conditions followed by